THREE ENT-KAURENE DITERPENES FROM VELLOZIA CAPUT-ARDEAE

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Key Word Index-Vellozia caput-ardeae; Velloziaceae; ent-kaurene diterpenes.

Abstract—Three new ent-kaurene diterpenes have been isolated from the roots and stem of Vellozia caput-ardeae. Their structures were elucidated by spectroscopic methods as ent-9β-hydroxy kaur-16-ene, ent-11α-hydroxy kaur-16ene and ent- 9β , 11α -dihydroxy kaur-16-ene.

INTRODUCTION

In previous papers [1, 2] we described two kaurene diterpenes and one seco-kaurene from Vellozia caputardeae. In the present communication we report the isolation and structural determination of three hydroxylated kaurenes from the same species.

RESULTS AND DISCUSSION

The molecular formulae of the isomeric alcohols 1a and **2a** $(C_{20}H_{32}O)$ and the diol **3a** $(C_{20}H_{32}O_2)$ were determined by high-resolution mass spectrometry. The ¹H NMR spectra of each of the three compounds contained resonances assigned to three tertiary methyl groups and an exocyclic methylene which suggested that they were tetracyclic diterpenes, probably of the kaurene class.

1a R = H, αOH

1b R=H, α OAc

1c R=0

2a

3a R = H, α OH 3b R = H, α OAc 3c R = 0

signals at δ 1.62 and 2.10 (both exchangeable with D_2O). This data determined that 3a was a diol. One of the hydroxyl groups was secondary, as shown by a double doublet at δ 3.96 (1H, J = 10 and 8 Hz) which moved downfield to 4.94 upon acetylation of 3a to 3b. The

The oxygen-containing functional group of 1a was shown to be a secondary hydroxyl group by IR $(v_{\text{max}}^{\text{KBr}} \text{ cm}^{-1}: 3450 \text{ and } 1060)$ and by the presence of a triple doublet at δ 4.28 (1H, J = 10, 8 and 8 Hz) in the ¹H NMR spectrum. This signal was shifted downfield to δ 5.37 upon acetylation of 1a to 1b. The multiplicity of this signal (ddd) indicated the vicinity of methine and methylene units. This limited the possible positions for the hydroxyl group to C-6 or C-11. The C-6 position was eliminated by analysis of the ¹³C NMR spectra (Table 1). In the spectrum for kaurene, the triplets for C-2, C-6 and C-11 are found near δ 20 while the triplets for C-7 and C-12 appear near δ 40 and 30, respectively [3]. As the C NMR spectrum of 1a exhibited only two triplets near δ 20 and absence of a triplet near 30, it was clear that C-11 was oxygenated. This was further corroborated by the spectroscopic properties of the ketone 1c derived from 1a. The ¹³C NMR signals for C-9 and C-12 showed the expected downfield shifts (Table 1) for 1c compared to 1a. Compound 1a was, thus, identified as ent-11 α hydroxykaur-16-ene.

Compound 2a was found to be a tertiary alcohol: IR $v_{\text{max}} \text{ cm}^{-1}$: 3500; ¹³C NMR singlet at δ 77.6, and the absence in the ¹H NMR spectrum of signals which could be assigned to a carbinolic proton.

Further evidence for the location of the hydroxyl group at C-9 was the dehydration product 2b of 2a, whose ¹H NMR spectrum exhibited a triplet at δ 5.14 (1H, J = 4 Hz)[4]. Comparison of the ¹³C NMR spectra of 2a and kaurene [3] confirmed the above deductions. The C-9 hydroxyl group in 2a caused an up-field shift in the resonances assigned to C-1, C-5, C-7 and C-15 through the strong y-effects within the cyclic system [5, 6]. The chemical shift of C-20, as expected, was unaltered in comparison to the model compound. This data also confirmed the stereochemistry assigned to 2a. Compound 2a was, therefore, identified as ent-9 β -hydroxy-kaur-16-

The ¹H NMR spectrum of compound 3a showed two

Table 1. 13C NMR chemical shifts of kaurene diterpend	oids*
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	1a	1 b	1c	2a	3a	3b	3e
C-1	42.3†	40.5	40.2	36.5	37.0	36.9	35.5
C-2	18.5	18.7	18.2	18.6	18.7	18.8	18.3
C-3	42.2†	41.4	41.6	41.6	42.2	41.0	41.4
C-4	33.6	33.5	33.4	33.3	33.8	33.9	33.7
C-5	56.9	56.3	55.5	48.0	50.1	48.3	46.6
C-6	20.2	20.2	20.0	20.1	20.1	20.0	20.1
C-7	42.0†	38.6	39.1	34.6	35.3	36.9	35.5
C-8	45.9	46.1	44.7	49.0	50.6	51.6	50.7
C-9	60.4	57.8	72.4	77.6	80.9	78.2	84.6
C-10	40.9	40.9	38.6	43.8	45.4	45.7	43.2
C-11	70.7	72.2	212.2	29.1	78.6	81.9	214.5
C-12	43.2	41.7	52.9	32.3	42.7	40.2†	52.2
C-13	39.6	43.1	43.1	42.3	40.7	41.9	43.2
C-14	41.8†	39.5	39.1	40.5	41.5	40.7†	42.8†
C-15	48.3	47.9	48.6	43.8	43.3	43.3	42.9†
C-16	154.8	153.9	152.2	155.0	154.4	154.1	152.3
C-17	103.1	104.1	106.6	102.8	102.9	103.7	106.6
C-18	34.1	33.9	33.6	33.7	34.2	33.5	33.4
C-19	21.8	21.7	21.7	21.8	22.0	21.8	21.9
C-20	18.7	18.8	18.6	19.2	19.3	19.3	20.5
(OAc)CO		170.3	_			173.7	
(OAc)Me	a man a specimen	20.2				21.6	

^{*}Values are in δ (ppm) downfield from TMS in CDCl₃.

presence of a tertiary hydroxyl group was confirmed by the ¹H NMR spectra of 3b and the ketone 3c, obtained from 3a by oxidation.

The position of the tertiary hydroxyl group was determined to be at C-9, from the ¹³C NMR chemical shifts of C-1, C-5, C-7 and C-15 in comparison with the spectrum of 2a.

The multiplicity of the signal assigned to the carbinolic proton, a double doublet, limited the location of the secondary hydroxyl group to C-1, C-7 or C-11. Positions C-1 and C-7 were eliminated by analysis of the ¹³C NMR spectra of compounds 3a-3c.

As regards the stereochemistry of two substances, 1a and 3a, the magnitude of the coupling constants for the carbinolic proton at C-11 in these substances and their derivatives (1b and 3b) made it obvious that this hydroxyl group was equatorial. Compound 3a was, thus, shown to be ent- 9β , 11α -dihydroxy-kaur-16-ene.

EXPERIMENTAL

Mps are uncorr. IR spectra were recorded on a Perkin-Elmer 137B. 1 H and 13 C NMR spectra at 100 and 25.2 MHz, respectively, and chemical shifts [δ (ppm)] measured from TMS as int. standard. Optical rotation was determined in CHCl₃ on a Perkin-Elmer 241.

Isolation of ent- 11α -hydroxy kaur-16-ene (1a) ent- 9β -hydroxy kaur-16-ene (2a) and ent- 9β , 11α -dihydroxy kaur-16-ene (3a). Chromatography of the hexane extract (51 g) of roots, stems and leaf sheaths of Vellozia caput-ardeae, collected in Diamantina, State of Minas Gerais, Brazil, yielded 1a, mp 106- 108° , 0.29° , of dry plant wt; IR v_{\max}^{KBr} cm⁻¹: 3450, 2940, 1650, 1060 and 880. ¹H NMR (100 MHz, CDCl₃): δ 0.84 (3H, s), 0.86 (3H, s), 1.22 (3H, s), 1.46 (1H, br s, exchangeable with D₂O), 4.28 (1H, ddd, J = 10, 8 and 8 Hz), 4.70 (1H, br s) and 4.82 (1H, br s). ¹³C NMR

(25.2 MHz, CDCl₃): δ 18.5 (t), 18.7 (q), 20.2 (t), 21.8 (q), 33.6 (s), 34.1 (q), 39.6 (d), 40.9 (s), 41.8 (t), 42.0 (t), 42.0 (t), 42.3 (t), 43.2 (t), 45.9 (s), 48.3 (t), 56.9 (d), 60.4 (d), 70.7 (d), 103.1 (t) and 154.8 (s). MS m/z (rel. int.): 228 [M] $^+$ (46), 273 (30), 270 (14), 255 (17), 163 (12), 146 (9), 137 (32), 123 (100), 109 (31), 107 (40), 91 (51), 69 (59), 55 (47) and 41 (86). Found m/z 288.2462, $C_{20}H_{32}O$ requires 288.2445.

Acetylation of 1a. Ac_2O (2 ml) was added to a soln of 1a (60 mg) in C_5H_5N (2 ml). The mixture was left for 12 hr at room temp., followed by the usual work-up. Recrystallization of the crude product from hexane and EtOAc yielded 1b (50 mg), mp 96–98°. IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 1725, 1235 and 890. ¹H NMR (100 MHz, CDCl₃): δ 0.84 (3H, s), 0.86 (3H, s), 1.26 (3H, s), 2.02 (3H, s), 4.76 (1H, br s), 4.86 (1H, br s) and 5.37 (1H, ddd, J = 12. 6 and 6 Hz). ¹³C NMR (25.2 MHz, CDCl₃): δ 18.7 (t), 18.8 (q), 20.2 (t), 20.2 (q), 21.7 (t), 33.5 (s), 33.9 (q), 38.6 (t), 39.5 (t), 40.5 (t), 40.9 (s), 41.4 (t), 41.7 (t), 43.1 (d), 46.1 (s), 47.9 (t), 56.3 (d), 57.8 (d), 72.2 (d), 104.1 (t), 153.9 (s) and 170.3 (s). MS m/z (rel. int.): 330 [M] $^+$ (2), 315 (4), 288 (18), 270 (23), 255 (28), 163 (8), 146 (30), 136 (52), 109 (29), 91 (41), 81 (43), 69 (60), 55 (50), 43 (100) and 41 (62).

Oxidation of 1a. Compound 1a (120 mg) was treated with pyridinium chlorochromate (140 mg) in dry CH₂Cl₂ (2 ml) at room temp. for 2 hr followed by the usual work-up, to yield 1c (100 mg), mp 112–113 ·· IR $\nu_{\rm max}^{\rm KBr}$ cm $^{-1}$: 2940, 1700, 1650 and 890. 1 H NMR (100 MHz, CDCl₃): δ 0.84 (3H, s), 0.90 (3H, s), 1.08 (3H, s), 2.46 (3H, m), 2.92 (1H, br s), 4.82 (1H, br s) and 4.92 (1H, br s) 13 C NMR (25.2 MHz, CDCl₃): δ 18.2 (t), 18.6 (q), 20.0 (t), 21.7 (q), 33.4 (s), 33.6 (q), 38.6 (s), 39.1 (t), 39.1 (t), 40.2 (t), 41.6 (t), 43.1 (d), 44.7 (s), 48.6 (t), 52.9 (t), 55.5 (d), 72.4 (d), 106.6 (t), 152.2 (s) and 212.2 (s). MS m/z (rel. int.): 286 [M] $^+$ (56), 271 (28), 149 (81), 123 (56), 105 (41), 91 (63), 55 (55) and 41 (100). CD (c 2.0 × 10 $^{-4}$ g/ml, cyclohexane): [θ]₃₅₀ 0, [θ]₃₃₃ + 2994, [θ]₃₂₁ + 6273, [θ]₃₁₀ + 6986, [θ]₃₀₀ + 5845, [θ]₂₆₀ 0.

ent-9 β -Hydroxy-kaurene (2a). Mp 84-85°, 0.3 % dry plant wt. IR $v_{\text{max}}^{\text{RB}}$ cm $^{-1}$: 3500, 2940, 1640 and 880. 1 H NMR (100 MHz,

[†]Signals may be reversed.

CDCl₃): δ 0.85 (3H, s), 0.90 (3H, s), 1.17 (3H, s), 3.50 (1H, s, exchangeable with D₂O) and 4.80 (2H, br s). ¹³C NMR (25.2 MHz, CDCl₃): δ 18.6 (t), 19.2 (q), 20.1 (t), 21.8 (q), 29.1 (t), 32.3 (t), 33.3 (s), 33.7 (q), 34.6 (t), 40.5 (t), 41.6 (t), 42.3 (d), 43.8 (s), 43.8 (t), 48.0 (d), 49.0 (s), 77.6 (s), 102.8 (t) and 155.0 (s). MS m/z (rel. int.): 288 [M] $^+$ (60), 270 (36), 255 (48), 163 (36), 146 (56), 136 (66), 123 (84), 109 (62), 91 (70), 69 (88), 55 (76) and 41 (100). Found m/z 288.2456, $C_{20}H_{32}O$ requires 288.2445.

Dehydration of 2a. To a soln of 2a (50 mg) in C_5H_5N (3 ml) was added POCl₃ (1 ml). The reaction mixture was allowed to stand overnight, and work-up gave a colourless oil of 2b (10 mg). ¹H NMR (60 MHz, CDCl₃): δ 0.84 (3H, s), 0.92 (3H, s), 1.06 (3H, s), 4.76 (1H, br s), 4.88 (1H, br s) and 5.14 (1H, t, J = 4 Hz). MS m/z (rel. int.): 270 [M]⁺ (14), 255 (55), 136 (24), 105 (32), 91 (53), 81 (26), 69 (54), 55 (63) and 41 (100).

ent-9 β ,11 α -Dihydroxy-kaurene (3a). Mp 162–163°, 0.36 % dry plant wt. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3450, 2940, 1655 and 880. ¹H NMR (100 MHz, CDCl₃): δ 0.86 (3H, s), 0.90 (3H, s), 1.34 (3H, s), 1.62 (1H, exchangeable with D₂O), 2.10 (1H, exchangeable with D₂O), 3.96 (1H, dd, J = 10 and 8 Hz), 4.72 (1H, br s) and 4.82 (1H, br s). ¹³C NMR (25.2 MHz, CDCl₃): δ 18.7 (t), 19.3 (q), 20.1 (t), 22.0 (q), 33.8 (s), 34.2 (q), 35.3 (t), 37.0 (t), 40.7 (d), 41.5 (t), 42.2 (t), 42.7 (t), 43.3 (t), 45.4 (s), 50.2 (d), 50.6 (s), 78.6 (d), 80.9 (s), 102.9 (t) and 154.4 (s). MS m/z (rel. int.): 304 [M] + (4), 286 (42), 271 (30), 163 (23), 151 (22), 136 (40), 123 (98), 109 (31), 91 (51), 81 (59), 69 (73), 55 (63) and 41 (100).

$$\left[\alpha\right]_{24^{\circ}}^{\lambda} = \frac{589 \quad 578 \quad 546 \quad 436 \quad 365}{-19.8 \quad -20.5 \quad -23.1 \quad -35 \quad -37} \text{ nm } (c \ 0.80).$$

Found m/z 304.2382, $C_{20}H_{32}O_2$ requires 304.2394.

Acetylation of (3a). Compound 3a was acetylated using the same procedure described for 2 above. Compound 3b was recrystallized from a mixture of hexane and EtOAc, mp 140–142°. IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3510, 2940, 1720, 1230 and 885. ¹H NMR (100 MHz, CDCl₃): δ 0.85 (3H, s), 0.88 (3H, s), 1.32 (3H, s), 2.08 (3H, s), 3.98 (1H, s, exchangeable with D₂O), 4.78 (1H, br s), 4.86 (1H, br s) and 4.94 (1H, dd, J = 12 and 8 Hz). ¹³C NMR (25.2 MHz, CDCl₃): δ 18.8 (t), 19.3 (q), 20.0 (t), 21.6 (q), 21.8 (q),

33.5 (*q*), 33.9 (*s*), 36.9 (*t*), 36.9 (*t*), 40.2 (*t*), 40.7 (*t*), 41.0 (*t*), 41.9 (*d*), 43.3 (*t*), 45.7 (*s*), 48.3 (*d*), 51.6 (*s*), 78.2 (*s*), 81.9 (*d*), 103.7 (*t*), 154.1 (*s*) and 173.7 (*s*). MS m/z (rel. int.): 346 [M] $^+$ (2), 331 (1), 286 (26), 271 (7), 253 (12), 215 (7), 189 (13), 163 (12), 149 (25), 136 (32), 123 (71), 109 (31), 91 (29), 81 (41), 69 (63), 55 (5), 43 (100) and 41 (63).

Oxidation of 3a. This was carried out using the same procedure described for 2a. Compound 3c, mp 144–145°, IR $v_{\text{max}}^{\text{KBT}}$ cm⁻¹: 3570, 2940, 1695, 1650 and 880. ¹H NMR (100 MHz, CDCl₃): δ 0.86 (3H, s), 0.92 (3H, s), 1.24 (3H, s), 3.82 (1H, s, exchangeable with D₂O) and 4.86 (2H, br d). ¹³C NMR (100 MHz, CDCl₃): δ 18.3 (t), 20.1 (t), 20.5 (q), 21.9 (q), 33.4 (q), 33.7 (s), 35.5 (t), 41.4 (t), 42.8 (t), 42.9 (t), 43.2 (s), 43.2 (d), 46.6 (d), 50.7 (s), 52.2 (t), 84.6 (s), 106.6 (t), 152.3 (s) and 214.5 (s). MS m/z (rel. int.): 302 [M]⁺ (100), 220 (15), 206 (9), 178 (15), 165 (35), 146 (29), 123 (26), 109 (15), 91 (20), 69 (26), 55 (21) and 41 (35).

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REFERENCES

- Pinto, A. C., Prado, S. K. and Pinchin, R. (1981) Phytochemistry 20, 520.
- Pinto, A. C., Prado, S. K., Filho, R. B., Hull, W. E., Neszmelyi, A. and Lukacs, G. (1982) Tetrahedron Letters 5267.
- 3. Hanson, J. R., Siverns, M., Piozzi, F. and Savona, G. (1976) J. Chem. Soc. Perkin Trans. 1, 114.
- Caballero, Y. and Walls, F. (1970) Bol. Inst. Quim. Univ. Nat. Auton. Mex. 79.
- Murakami, T., Iida, H., Tanaka, N., Saiki, Y., Chen, C. M. and Iitaka, Y. (1981) Chem. Pharm. Bull. 29, 657.
- Tanaka, N., Murakami, T., Saiki, Y., Cheng, C. M., and Iitaka, Y. (1981) Chem. Pharm. Bull 29, 663.